

# Separation of Monoclonal Antibodies Using TSK-GEL HPLC Columns

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## 1. Introduction

In recent years, advances in biotechnology have resulted in increased use of monoclonal antibodies (MAbs) as the main constituent of therapeutic drugs and as diagnostic agents. Because of this increased commercial application, it is important for the bioanalytical chemist to have access to the tools for rapid and simple purification of MAbs. There are many published reports about MAb purification by high-performance liquid chromatography (HPLC) and FPLC (fast protein liquid chromatography); a select group of references is included at the end of this Separation Report. This document describes the analysis and isolation of MAbs by HPLC using TSK-GEL columns. Several application examples are discussed.

## 2. Separation Mode and Purification Method

Table 1 shows the HPLC separation modes used in the analysis and isolation of MAbs, together with their main features. Ion exchange (IEC), hydrophobic interaction (HIC) and affinity (AFC) have large sample capacity and high selectivity; as such, they are often used in MAb purification. IEC is said to be used in almost 75% of the MAb purifications, often following ammonium sulfate fractionation (HIC). Though gel filtration chromatography (GFC) is inferior with respect to resolution or sample load, it is often used as the final purification method and as a purity check for MAb preparations since this method constitutes a simple step for the removal of dimers and higher molecular weight aggregates.

**Table-1 Features of HPLC separation modes for monoclonal antibody purification**

Separation mode	Separation mechanism	TSK-GEL	Feature
Gel filtration (size exclusion) chromatography (GFC, SEC)	Molecular size	TSK-GEL G2000SW <sub>XL</sub> TSK-GEL G3000SW <sub>XL</sub> TSK-GEL G4000SW <sub>XL</sub>	Though resolution and sample load are inferior to other separation modes, it allows simple removal of dimers and aggregates, causes of MAb antigenicity. It can also be used in final purification or as a purity check. Separation of Fab and IgM (TSK-GEL G4000SW <sub>XL</sub> ) is also possible.
Ion exchange chromatography (IEC)	Ionic interaction	TSK-GEL DEAE-5PW TSK-GEL SP-5PW TSK-GEL CM-5PW  TSK-GEL DEAE-NPR TSK-GEL SP-NPR	Resolution is high and the sample load is greater. Anion exchange plays an important role for the separation of MAb sub-classes and impurities. Cation exchange allows simple purification by step-wise elution because many of the impurities are not retained on the column.  The non-porous resin (NPR) column can be used for rapid analysis and fractionation of trace amounts of MAb ( $\mu\text{g}$ to ng); purity check of MAb fractions is possible instead of electrophoresis.
Hydrophobic interaction chromatography (HIC)	Hydrophobicity	TSK-GEL Ether-5PW TSK-GEL Phenyl-5PW TSK-GEL Butyl-NPR	Resolution is high and the sample load is good. TSKgel Ether-5PW, particularly, has high recovery of hydrophobic MAbs. HIC allows simple purification by step-wise elution. TSKgel Butyl-NPR is ideal for micro-fractionation.
Affinity chromatography (AFC)	Bioaffinity	TSK-GEL Chelate-5PW TSK-GEL Tresyl-5PW	Immobilized metal chelate affinity chromatography (IMAC). Activated affinity support. Ligands such as antigen are immobilized to absorb and purify MAb.

In general, it is said that using a single HPLC mode (with the exception of GFC) provides a MAb purity of about 95%. Thus, high purity preparations of MAb for use in diagnostic applications can be obtained using a single HPLC column. However, a purity of 99% and higher is required when MAbs are used for medicinal purposes. This level of purity can be achieved using a single Protein A affinity step, which is based on antibody-antigen interaction. A minimum of two different separation modes are required when less specific HPLC separation modes are applied, such as IEC-HIC or IEC-GFC.

Furthermore, recent demands for preparation of high purity MAb solutions (approx. 99.9% purity or higher) require three different separation modes, for instance, HIC-IEC-GFC or AFC-IEC-GFC. In addition, it is necessary to select the proper elution method, whether linear elution, gradient elution or step-wise gradient elution, depending on the purity required, the time allowed for purification and the cost of each process of preparing the MAb preparation. Table 2 lists the literature in which various TSK-GEL products were used for MAb purification along with a brief summary about each mode. While most methods consist of IEC and GFC, reports are included based on HIC and AFC as well.

**Table-2 Key literature featuring TSK-GEL columns for the separation of monoclonal antibodies**

Literature No.	Column	Sample	Antibody type	Outline
1	TSK-GEL G3000SW	Ascites, cultured supernatant	IgG <sub>1</sub> , F(ab) <sub>2</sub>	Review article on MAb against class I, MHC antigen
2	TSK-GEL G3000SW	Ascites	IgG <sub>1</sub> , F(ab) <sub>2</sub>	Separation of IgG <sub>1</sub> and F(ab) <sub>2</sub>
3	TSK-GEL G3000SW	Ascites	IgG <sub>1</sub> , IgG <sub>2b</sub> , IgG <sub>3</sub>	Final purification of MAb purified by IEC Recovery is 85% and higher
4	TSK-GEL DEAE-5PW TSK-GEL G3000SW	Ascites, cultured supernatant	IgG <sub>1</sub> , IgG <sub>2a</sub> to the total of 8 species	1 step IEC purification of MAb by sophisticated eluent Recovery is 95%
5	TSK-GEL G3000SW	Ascites	IgG <sub>1</sub> , IgG <sub>2a</sub> , IgG <sub>2b</sub>	Purity check on MAb purified by IEC
6	TSK-GEL DEAE-5PW	Ascites	IgG <sub>1</sub> , IgG <sub>2a</sub> , IgG <sub>2b</sub>	1 step IEC purification of MAb by sophisticated eluent Recovery is 92% and higher
7	TSK-GEL Phenyl-5PW	Ascites, cultured supernatant	IgG	Examination of separation by IEC and HIC IEC and HIC were found to provide good separation
8	Toyopearl DEAE-650M Toyopearl CM-650M	Cultured supernatant	IgG <sub>2b</sub>	Rapid separation of sample; simple removal of phenol red indicator. Recovery of activity is 90% and higher
9	TSK-GEL SP-5PW TSK-GEL G4000SW	Ascites	IgG <sub>1</sub> , IgM	Simple purification by IEC, automatic control by IEC-GFC connection. Recovery of activity is 80% and higher
10	TSK-GEL SP-5PW	Cultured supernatant	IgG	Step-wise gradient elution of pretreated sample (100mL) Purity 99%
11	TSK-GEL Chelate-5PW	Ascites	IgG	Application of IMAC, purity increased 10-fold in single step
12	TSK-GEL DEAE-5PW TSK-GEL G3000SW	Ascites	IgG <sub>1</sub> , IgG <sub>2a</sub> , IgG <sub>2b</sub> , IgM	MAb purification on preparative columns using a sophisticated eluent. Recovery is 89% and higher
13	TSK-GEL DEAE-5PW TSK-GEL G3000SW	Ascites	IgG <sub>1</sub>	Comparative examination of purification by IEC and AFC on preparative columns. Purity 99.2% and recovery of activity 72%
14	TSK-GEL DEAE-5PW	Ascites	IgM (2 species)	Purification of IgM to high purity in 1 step using simultaneous gradient of salt and pH
15	TSK-GEL Ether-5PW TSK-GEL G3000SW <sub>XL</sub> TSK-GEL DEAE-NPR	Ascites	IgG <sub>1</sub>	Examination of MAb separation conditions by HIC, simple purification in step-wise elution

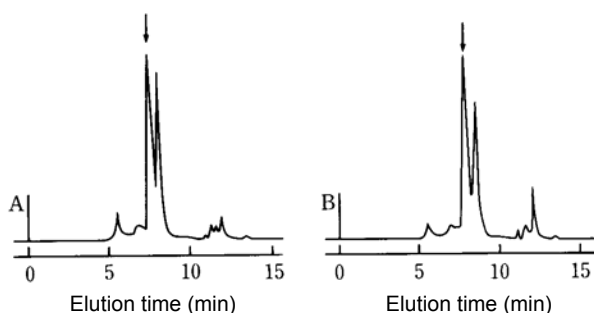
### 3. Applications

#### 3.1 Gel Filtration Chromatography (GFC, SEC)

GFC is inferior to other chromatographic modes for the separation of MAb in terms of resolution and sample load. IgG is often expressed in mouse ascites fluid in the presence of large amounts of albumin. When injecting crude samples into a GFC column, it is often not possible to obtain satisfactory resolution between MAb (IgG) and albumin. Thus GFC is more commonly used for separating partially purified samples rather than for direct MAb purification from crude ascites samples.

Figure 1 shows an example of MAb (IgG<sub>1</sub>) separation by GFC from mouse ascites fluid. Although it was found that separation of IgG and albumin could be improved somewhat by changing the eluent pH and salt species (pH 5.0, 0.1mol/L Na<sub>2</sub>SO<sub>4</sub>), the resolution would not be sufficient for most applications. For a detailed examination of this separation, see Separation Report No. 62.

To isolate IgM, TSK-GEL G4000SW<sub>XL</sub> is best suited since the larger pore size enables separation of larger proteins.

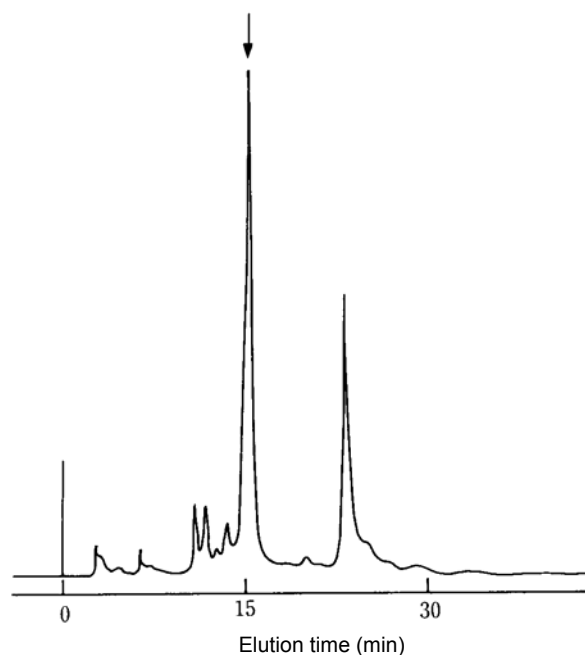


**Figure 1 Separation of monoclonal antibody by GFC**

Column: TSKgel G3000SW<sub>XL</sub>, 7.8mm ID × 30cm  
Eluent: A: 50mmol/L sodium phosphate buffer (pH 6.7) + 0.3mol/L NaCl  
B: 50mmol/L sodium phosphate buffer (pH 5.0) + 0.1mol/L Na<sub>2</sub>SO<sub>4</sub>  
Flow rate: 1.0mL/min  
Temperature: 25°C  
Detection: UV@280nm  
Sample: Anti-human proline hydroxylase (IgG<sub>1</sub>), diluted solution of mouse ascites (50µl)

#### 3.2 Ion-Exchange Chromatography (IEC)

Ion Exchange Chromatography steps either consist of an anion exchange packing material with DEAE (diethylaminoethyl) functional groups or a cation exchange packing material with SP (sulfopropyl) or CM (carboxymethyl) functional groups. In general, DEAE anion exchangers are used as the last step in MAb purifications. DEAE is also very suitable for the separation of IgG subclasses. In both cases a linear gradient method is applied. Best separations are obtained at pH 8 or higher. Figures 2 and 3 show the separation of MAb (IgG<sub>1</sub>) from mouse ascites fluid and cell culture supernatant on TSK-GEL DEAE-5PW. In the case of mouse ascites fluid, shown in Figure 2, IgG<sub>1</sub> elutes in about 15 minutes, well separated from the impurities that elute before or after IgG<sub>1</sub> such as transferrin (about 11 minutes) and albumin (about 22 minutes). However, when injecting cell culture supernatant, as shown in Figure 3, MAb could not be detected because it was present at trace levels.



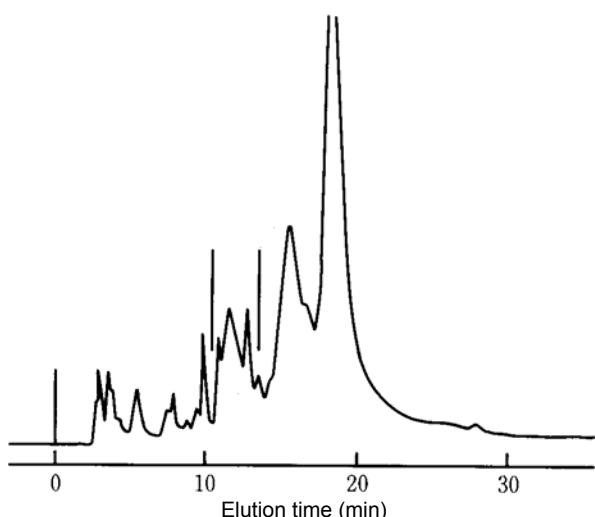
**Figure 2 Separation of monoclonal antibody by IEC**

Column: TSKgel DEAE-5PW, 7.5mm ID × 7.5cm  
Eluent: A: 20mmol/L Tris-HCl (pH 8.5)  
B: A + 0.5mol/L NaCl  
A → B linear gradient (60 min.)  
Flow rate: 1.0mL/min  
Temperature: 25°C  
Detection: UV@280nm  
Sample: Anti-human albumin (IgG<sub>1</sub>), diluted solution of mouse ascites (168µg in 40µl)

In contrast to anion exchange, with a cation exchanger such as SP or CM, sample throughput is higher because albumin and other abundant proteins pass through the column instead of being absorbed at the usual eluent pH. It is also possible to purify MABs simply by step-wise gradient elution. Figure 4 shows a chromatogram in which MAb (IgG<sub>1</sub>) is separated from cell culture supernatant on a TSKgel SP-5PW column. Clearly, the elution pattern and the selectivity differ as a function of the buffer in the eluent. In this case, better results are obtained with acetate rather than citrate buffer.

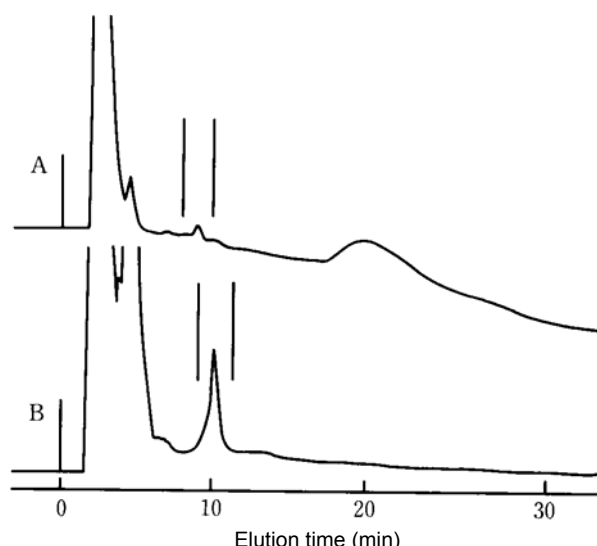
Though non-porous packing materials are not suitable to purify large quantities of monoclonal antibodies, they are the right column type to conduct a quick purity check of a MAB preparation since the high efficiency of NPR columns allow for fast separations of trace sample amounts.

Figure 5 demonstrates that a sample of mouse ascites fluid can be separated on a TSKgel DEAE-NPR column with similar resolution as that obtained on a TSKgel DEAE-5PW column (see Figure 2). Non-porous resin columns such as TSKgel DEAE-NPR are especially useful to check sample purity or to monitor expression levels, as they allow very short analysis times. Finally, although sample capacity is limited, columns packed with non-porous resin can be used to isolate nanogram amounts of MAB in a sample (see Separation Report No.65 for details).



**Figure 3 Separation of monoclonal antibody by IEC**

Column: TSKgel DEAE-5PW, 7.5mm ID × 7.5cm  
 Eluent: A: 20mmol/L Tris-HCl (pH 8.8)  
 B: A + 0.5mol/L NaCl  
 A → B linear gradient (30 min.)  
 Flow rate: 1.0mL/min  
 Temperature: 25°C  
 Detection: UV@280nm  
 Sample: Anti-HLA-A, B, C (IgG<sub>1</sub>),  
 NS-1 culture supernatant

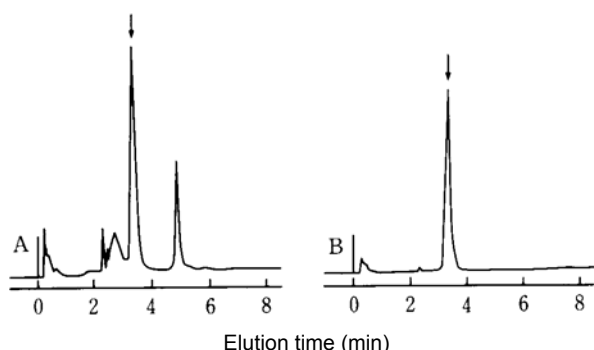


**Figure 4 Separation of monoclonal antibody on IEC**

Column: TSKgel SP-5PW, 7.5mm ID × 7.5cm  
 Eluent: A: a: 20mmol/L citrate buffer (pH 5.7)  
 b: a + 0.5mol/L NaCl  
 a → b linear gradient (30 min.)  
 B: c: 20mmol/L acetate buffer (pH 5.7)  
 d: c + 0.5mol/L NaCl  
 c → d linear gradient (30 min.)  
 Flow rate: 1.0mL/min  
 Temperature: 25°C  
 Detection: UV@280nm  
 Sample: Anti-HLA-A, B, C (IgG<sub>1</sub>),  
 NS-1 culture supernatant (500µg in 500µl)

### 3.3 Hydrophobic Interaction Chromatography (HIC)

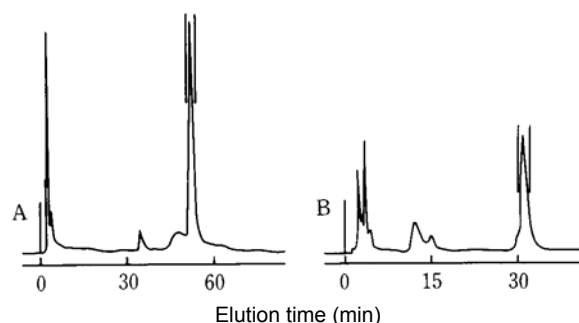
HIC is widely used as a method of protein purification and it is now often one of the steps in MAb purification. Traditionally, TSK-GEL Phenyl-5PW has mainly been used for the separation of proteins by HIC, while TSK-GEL Ether-5PW is better suited for the separation and recovery of such hydrophobic proteins as membrane proteins and proteins with molecular weights of 100,000Da or higher.



**Figure 5** Purity check on monoclonal antibody fraction by IEC

Column: TSKgel DEAE-NPR, 4.6mm ID × 3.5cm  
 Eluent: a: 50mmol/L Tris-HCl (pH 8.5)  
 b: a + 0.5mol/L NaCl  
 a → b linear gradient (10 min.)  
 Flow rate: 1.5mL/min  
 Temperature: 25°C  
 Detection: UV@280nm  
 Sample: Anti-fowl 14K lectin (IgG<sub>1</sub>)  
 A: Mouse ascites (× 4, 5μl)  
 B: TSKgel HA-1000, MAb fraction (100μl)

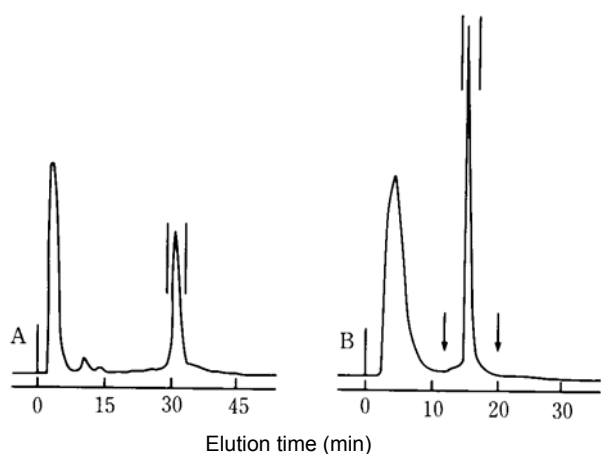
Figure 6 shows the separation of mouse ascites on TSK-GEL Phenyl-5PW and Ether-5PW columns, respectively. On the TSKgel Phenyl-5PW column the MAb (IgG<sub>1</sub>) peak overlaps with the albumin peak at about 47 minutes. In contrast, on the TSKgel Ether-5PW column the MAb peak is well separated from the albumin peak, which elutes at about 13 minutes. In addition, MAb elutes faster from the TSKgel Ether-5PW column than TSKgel Phenyl-5PW when using the same separation conditions due to the difference in hydrophobicity of the packing materials. As shown in Figure 6, it is evident that in this study TSK-GEL Ether-5PW is the better choice for MAb separation by HIC.



**Figure 6** Separation of monoclonal antibody by HIC

Column: A: TSKgel Phenyl-5PW, 7.5mm ID × 7.5cm  
 B: TSKgel Ether-5PW, 7.5mm ID × 7.5cm  
 Eluent: a: 0.1mol/L sodium phosphate buffer (pH 7.0) + 1.5mol/L ammonium sulfate  
 b: 0.1mol/L sodium phosphate buffer (pH 7.0)  
 a → b linear gradient (60 min.)  
 Flow rate: 1.0mL/min  
 Temperature: 25°C  
 Detection: UV@280nm  
 Sample: Anti-fowl 14K lectin (IgG<sub>1</sub>)  
 Mouse ascites (1.5mg in 100μl)

Figure 7 shows that a large sample volume can be injected on the TSKgel Ether-5PW column. 3mL of mouse ascites (2-fold dilution, 1.0mol/L ammonium sulfate) was directly injected on the column and eluted with a linear gradient or a step-wise gradient elution. A chromatogram nearly identical to Figure 6 was obtained for linear gradient elution (data not shown). With step-wise gradient elution, most of the impurities pass through the column, while only MAb is absorbed at the starting ammonium sulfate concentration of 1.0mol/L. The absorbed MAb was eluted with 0.5mol/L ammonium sulfate. Clearly, step-wise gradient elution provided a simple separation of this large sample volume.



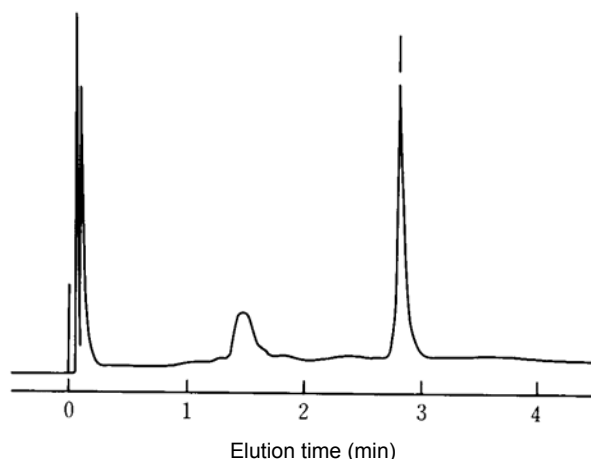
**Figure 7 Separation of large amount of monoclonal antibody by HIC**

Column: TSKgel Ether-5PW, 7.5mm ID × 7.5cm  
 Eluent: A: Same as Figure 6  
 B: a: 0.1mol/L sodium phosphate buffer (pH 7.0) + 1.0mol/L ammonium sulfate  
 b: 0.1mol/L sodium phosphate buffer (pH 7.0)  
 a → a/b (50/50) (12 min.) → b (20 min.) step-wise gradient  
 Flow rate: 1.0mL/min  
 Temperature: 25°C  
 Detection: UV@280nm  
 Sample: Anti-fowl 14K lectin (IgG<sub>1</sub>),  
 Mouse ascites (90mg in 3.0mL)

Moreover, if one wishes to purify a large quantity of MAb, it is also possible to employ packing materials developed for low pressure liquid chromatography (Toyopearl resins) instead of using an HPLC column.

Toyopearl Ether-650M has a much larger particle size (~65µm) than TSK-GEL Ether-5PW (10µm) and is thus less efficient, although its selectivity is similar to that for TSK-GEL Ether-5PW. Please consult our website or request a copy of the Process Chromatography catalog for more information about the relationship between TSK-GEL columns and Toyopearl resins.

Similar to IEC, a non-porous resin column is available in the TSK-GEL line of HIC columns. Using TSK-GEL Butyl-NPR, MAb purity check or micro-purification can be rapidly achieved. Figure 8 shows the separation of mouse ascites on a TSKgel Butyl-NPR column. The separation was completed in about 3 minutes and it is clear that MAb is well separated from the impurities.

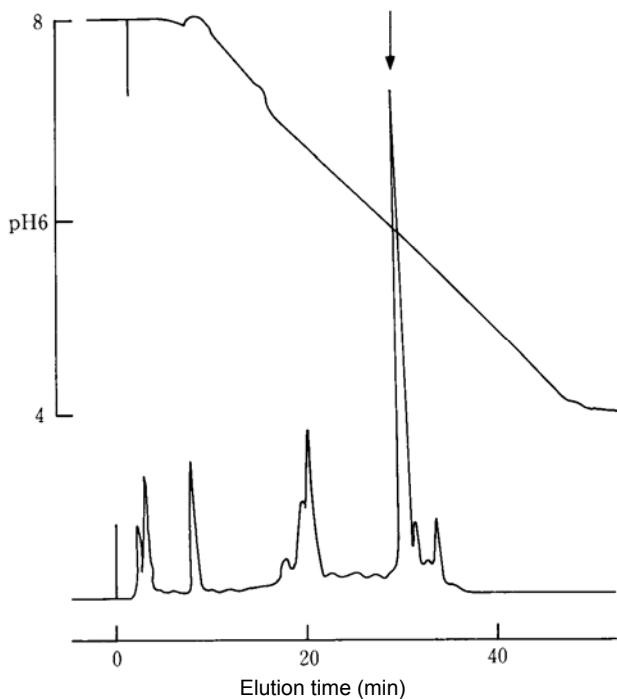


**Figure 8 Separation of monoclonal antibody by HIC**

Column: TSKgel Butyl-NPR, 4.6mm ID × 3.5cm  
 Eluent: A: 0.1mol/L sodium phosphate buffer (pH 7.0) + 1.5mol/L ammonium sulfate  
 B: 0.1mol/L sodium phosphate buffer (pH 7.0)  
 A → B linear gradient (10 min.)  
 Flow rate: 1.0mL/min  
 Temperature: 25°C  
 Detection: UV@280nm  
 Sample: Anti-fowl 14K lectin (IgG<sub>1</sub>)  
 Mouse ascites (38µg in 2.5µl)

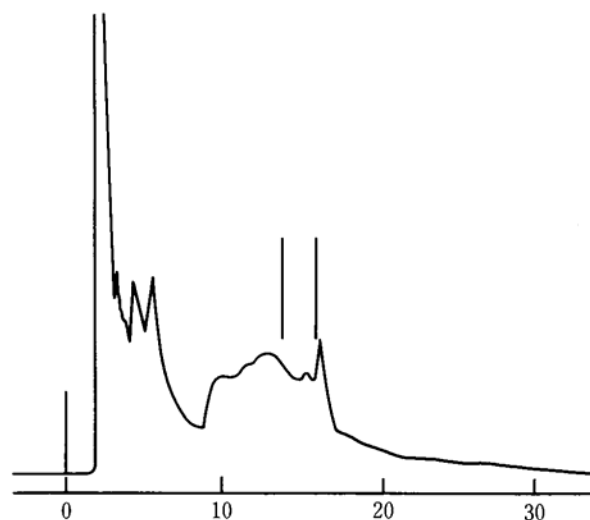
### 3.4 Affinity Chromatography (AFC)

AFC using Protein A is often employed for MAb separation, however, it is also possible to separate monoclonal antibodies using Immobilized Metal Ion Affinity Chromatography (IMAC). Figures 10 and 11 demonstrate MAb purification from two matrices, mouse ascites and culture supernatant, on a TSKgel Chelate-5PW column loaded with  $Zn^{2+}$  ion. Figure 11 shows that MAb ( $IgG_1$ ) is eluted in about 32 minutes by pH gradient elution and is well separated from other impurities.



**Figure 10 Separation of monoclonal antibody by AFC**

Column: TSKgel Chelate-5PW Glass ( $Zn^{2+}$ ),  
8mm ID  $\times$  7.5cm  
Eluent: A: 20mmol/L HEPES-MES-acetate buffer  
(pH 8.0) + 0.5mol/L NaCl  
B: 20mmol/L HEPES-MES-acetate buffer  
(pH 4.0) + 0.5mol/L NaCl  
A  $\rightarrow$  B linear gradient (40 min.)  
Flow rate: 1.0mL/min  
Temperature: 25°C  
Detection: UV@280nm  
Sample: Anti-fowl 14K lectin ( $IgG_1$ ),  
Mouse ascites (50 $\mu$ l)



**Figure 11 Separation of monoclonal antibody by AFC**

Column: TSKgel Chelate-5PW ( $Zn^{2+}$ ),  
7.5mm ID  $\times$  7.5cm  
Eluent: A: 20mmol/L Tris-HCl (pH 8.0) + 0.5mol/L  
NaCl  
B: A + 200mmol/L glycine  
A  $\rightarrow$  B linear gradient (30 min.)  
Flow rate: 1.0mL/min  
Temperature: 25°C  
Detection: UV@280nm  
Sample: Anti-HLA-A, B, C ( $IgG_1$ ),  
NS-1 cultured supernatant

## 4. Conclusion

As discussed in this document, there are several different separation modes for the purification of monoclonal antibodies and each separation mode has its advantages. Therefore, it is important in the purification of monoclonal antibodies to select the most appropriate separation mode and column with consideration of various factors for purification, such as resolution, sample load, operability, cost, and purity.

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